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Seagrass-derived fungi as a source of bioactive compounds

Five new eudesmane-type sesquiterpenes – thomimarines A-E (1-5) and 12 new polyketides with "decalin" moiety – zosteropenillines A-L (6-17) were isolated from the ethylacetate extract of the fungi Penicillium thomii associated with the seagrass Zostera marina. Their structures were established based on spectroscopic methods. The absolute configurations of 1-5 and 7-10 were determined by time-dependent density functional theory (TD-DFT) calculations of ECD spectra. The absolute configuration of zosteropenilline A (6) was determined by a combination of the modified Mosher's method, X-ray analysis, and NOESY data. The effect of selected compounds on the viability of human drug-resistant prostate cancer cells PC3 as well as on autophagy in these cancer cells and inhibitory effects of selected compounds on NO production in LPS-induced RAW 264.7 murine macrophages were evaluated.

Key words: Penicillium thomii, Zostera marina, NMR, X-ray, ECD spectra, TD-DFT, Mosher's method, autophagy, NO, sesquiterpenes, polyketides.

Marine-derived fungi are a prolific source of new secondary metabolites many of which are biologically active [4-6]. As part of our ongoing search for structurally novel and bioactive metabolites we have isolated 17 new compounds from two fungi *Penicillium thomii*, associated with seagrass *Zostera marina* (Troitsa bay, Sea of Japan), including five new eudesmanetype sesquiterpenes – thomimarines A-E (1-5) [1, 2] (Figure 1) from the *P. thomii* KMM 4667 and 12 new polyketides with "decalin" moiety – zosteropenillines A-L (6-17) (Figure 2) from the *P. thomii* KMM 4674 [3] (Figure 1). Their structures were established based on spectroscopic methods. The absolute configurations of thomimarines A-E (1-5) and zosteropenillines B-D (7-9) were determined by time-dependent density functional theory (TD-DFT) calculations of ECD spectra. The absolute configuration of zosteropenilline A (6) was determined by a combination of the modified Mosher's method, X-ray analysis and NOESY data (Figure 3).



Figure 1. Chemical structures of thomimarines A-E (1-5)

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It was found that compounds 1, 4, 5, 6, 13 and 15 at concentration of 10.0 μ M induced a moderate down-regulation of NO production in macrophages stimulated with LPS. NO level in these cells was decreased by 24.9%±0.9, 20.9%±5.7, 22.5±5.1%, 27.7%±1.8, 20.6%±1.2 and 22.3%±3.8, respectively, compare to control cells pretreated with LPS.



Figure 2. Chemical structures of zosteropenillines A-L (6-17)

The most effective substance was compound **2** exhibited the maximal pronounced inhibition of NO formation in LPS-stimulated RAW 264.7 cells by $43.4\%\pm1.5$. The effect of zosteropenillines **6-8**, **12**, **13**, **15** and **16** on the viability of human drug-resistant prostate cancer cells PC3 as well as on autophagy in these cancer cells was examined. The results suggest that the investigated compounds are able to inhibit autophagy at non-cytotoxic concentrations and may sensibilize human cancer cells to cytotoxic anticancer drugs.



Figure 3. (A) Key HMBC and COSY correlations of 6; (B) $\Delta\delta(\delta_{s} - \delta_{R})$ values (in ppm) for the (S)- and (R)-MPTA esters of 6; (C) Crystal structure of 6

REFERENCES:

1. Afiyatullov S.S., Leshchenko E.V., Sobolevskaya M.P., Denisenko V.A., Kirichuk N.N., Khudyakova Y.V., Hoai T.P.T., Dmitrenok P.S., Menchinskaya E.S., Pislyagin E.A., Berdyshev D.V. New eudesmane sesquiterpenes from the marine-derived fungus *Penicillium thomii* // Phytochem. Lett. 2015. Vol. 14. P. 209 - 214.

2. Afiyatullov S.S., Leshchenko E.V., Sobolevskaya M.P., Antonov A.S., Denisenko V.A., Popov R.S., Khudyakova Y.V., Kirichuk N.N., Kuz'mich A.S., Pislyagin E.A., Kim N.Y., Berdyshev D.V. New Thomimarine E from Marine Isolate of the Fungus *Penicillium thomii* // Chem. Nat. Compd. 2017. Vol. 53, N. 2. P. 290 - 294.

3. Afiyatullov S.S., Leshchenko E.V., Berdyshev D.V., Sobolevskaya M.P., Antonov A.S., Denisenko V.A., Popov R.S., Pivkin M.V., Udovenko A.A., Pislyagin E.A., Von Amsberg G., Dyshlovoy S.A. Zosteropenillines: Polyketides

from the marine-derived fungus *Penicillium thomii* // Mar. Drugs 2017. Vol. 15, N. 2. P. 1-17. DOI: 10.3390/md15020046.
4. Blunt J.W., Copp B.R., Keyzers R.A., Munro M.H.G., Prinsep M.R. Marine natural products // Nat. Prod. Rep. 2016. Vol. 33, N. 3. P. 382 - 431.

5. Jin L., Quan C., Hou X., Fan S. - Potential Pharmacological Resources: Natural Bioactive Compounds from Marine-Derived Fungi // Mar. Drugs 2016. Vol. 14, N. 4. DOI: 10.3390/md14040076.

6. Rateb M.E., Ebel R. Secondary metabolites of fungi from marine habitats // Nat. Prod. Rep. 2011. Vol. 28, N. 2. P. 290 - 344.